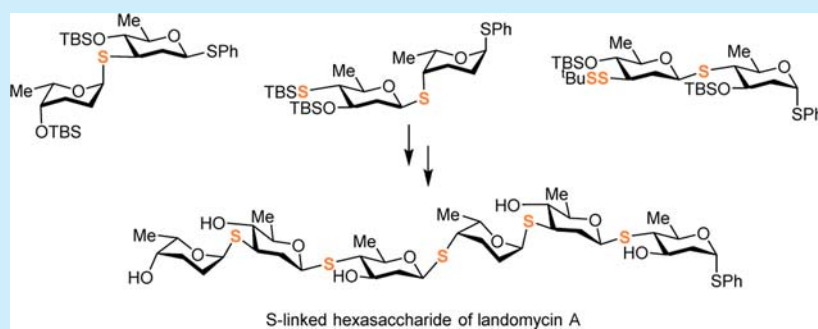


Stereochemical Synthesis of S-Linked Hexasaccharide of Landomycin A via Umpolung S-Glycosylation

Kedar N. Baryal and Jianglong Zhu*

Department of Chemistry and Biochemistry and School of Green Chemistry and Engineering, The University of Toledo, 2801 West Bancroft Street, Toledo, Ohio 43606, United States

Supporting Information



ABSTRACT: Stereoselective synthesis of carbohydrate mimics resistant toward acid-mediated or enzymatic hydrolysis is chemically challenging and biologically interesting. In this Letter, the first stereoselective synthesis of a “non-hydrolyzable” S-linked hexasaccharide of antitumor antibiotic landomycin A is described. This synthesis was accomplished through the utilization of our recently developed umpolung reactivity-based S-glycosylation–sulfenylation of stereochemically defined glycosyl lithium species with asymmetric sugar-derived disulfides.

The landomycins are a family of angucycline antitumor antibiotics isolated from *Streptomyces cyanogenus*.¹ In particular, landomycin A (cf. 1, Figure 1) possesses a broad

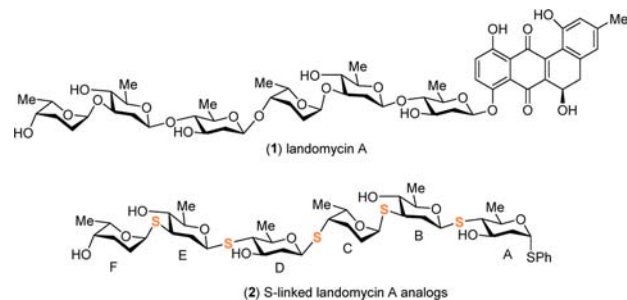


Figure 1. Landomycin A (1) and its associated S-linked hexasaccharide (2).

spectrum antitumor activity against 60 cancer cell lines.^{1a,2} Structurally, landomycin A contains an angular tetracyclic core as well as a hexasaccharide subunit consisting of two repeat units of trisaccharide (α -L-rhodinose-(1 \rightarrow 3)- β -D-olivose-(1 \rightarrow 4)- β -D-olivose). Although it is known that landomycin A inhibits DNA synthesis and G1/S cell cycle progression,³ the specific mechanism of action on cancer cells has not yet been determined. SAR studies suggested that cytotoxic activity of the landomycins depends on the length of the glycan subunit. In this respect, landomycin A with the longest sugar chain was the most potent congener, while landomycins B, D, E, I, and J with

shorter sugar chains showed diminished cytotoxic activities.^{1,3} The initial structure of landomycin A proposed by Rohr^{1a} in 1990 was revised in 1994⁴ and later confirmed through synthetic studies by Roush⁵ and a recent total synthesis from the Yu group.⁶ The hexasaccharide subunit of landomycin A has also been previously synthesized by three independent groups including Sulikowski,⁷ Roush,⁸ and Yu.⁹ In addition, the repeating trisaccharide has been prepared by Kirschning¹⁰ and O'Doherty.¹¹ Furthermore, stereoselective synthesis of a combinatorial library of 16 deoxyhexasaccharides related to the landomycin A sugar moiety has also been recently reported.¹²

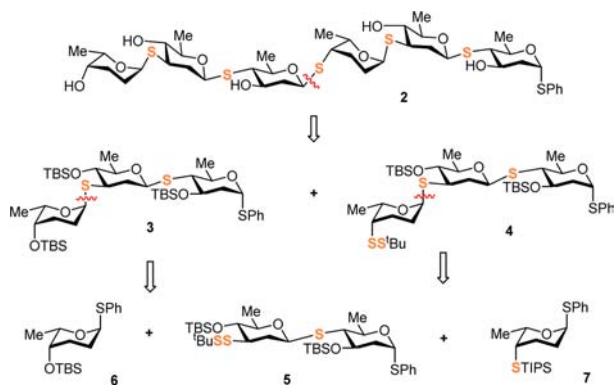
It is well-known that 2-deoxy glycosidic linkages are susceptible to acid-mediated or enzymatic hydrolysis. In order to access carbohydrate mimics resistant toward hydrolysis, recently our laboratory has developed an umpolung reactivity-based S-glycosylation¹³ for the stereoselective preparation of S-linked 2-deoxy glycosides¹⁴ (2-deoxy thioglycosides).¹⁵ Herein, we describe the first total synthesis of the S-linked hexasaccharide of landomycin A (cf. 2, Figure 1) employing the aforementioned umpolung reactivity-based S-glycosylation.

Our initial retrosynthesis of S-linked hexasaccharide 2 is depicted in Scheme 1. In this, 2 would be obtained by 3 + 3 coupling of trisaccharide donor 3 (fragment DEF) with trisaccharide-derived asymmetric disulfide acceptor 4 (fragment

Received: July 30, 2015

Published: September 3, 2015

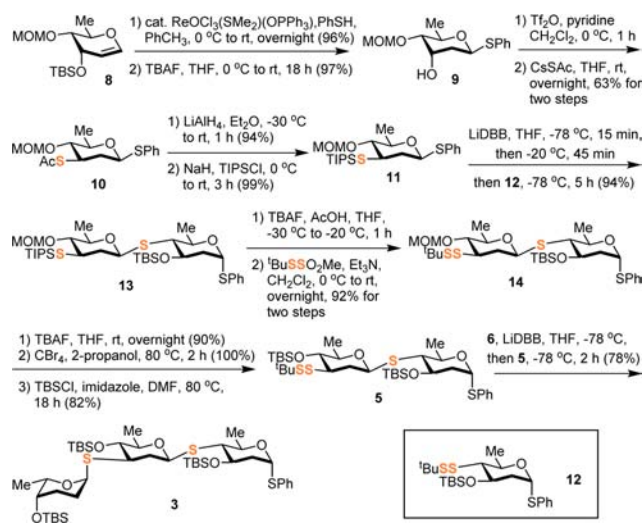
Scheme 1. Initial Retrosynthesis of S-Linked Hexasaccharide of Landomycin A (2)



ABC) via our previously reported umpolung S-glycosylation.¹³ In turn, donor 3 would be obtained from disaccharide-derived disulfide 5 and donor 6,^{13a} while acceptor 4 would be accessed from disaccharide-derived disulfide 5 and donor 7.

In order to synthesize trisaccharide donor 3, known protected 6-deoxy-D-allal 8¹⁶ underwent Re(V)-catalyzed thioglycosylation^{14b} with thiophenol followed by desilylation to furnish compound 9 (Scheme 2). Triflation of the C3-

Scheme 2. Synthesis of Trisaccharide Donor (3)

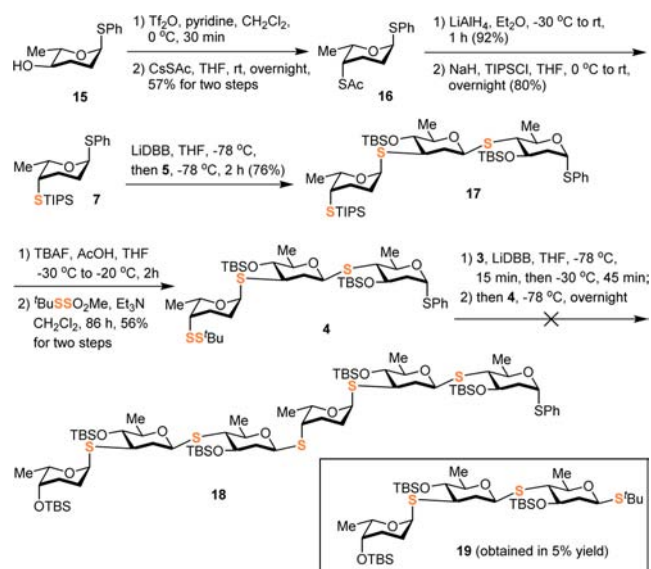


alcohol of 9 followed by S_N2 substitution with cesium thioacetate gave desired product 10. Reductive removal of the S-acetyl group afforded the corresponding free thiol which was subsequently protected as triisopropylsilyl thioether 11. Next, lithium 4,4'-di-*tert*-butylbiphenyl (LiDBB)-mediated reductive lithiation of the glycosyl phenylsulfide of 11 followed by anomerization (−20 °C)¹³ afforded the corresponding equatorial lithium which reacted with asymmetric disulfide acceptor 12^{13a} to give rise to the desired β-S-linked disaccharide 13 in 94% yield. Selective deprotection of triisopropylsilyl thioether of 13 in the presence of TBS ether led to the formation of the free thiol which reacted with *S*-*tert*-butylmethanethiosulfonate to afford asymmetric disulfide 14. Despite several attempts, the selective deprotection of methoxymethyl (MOM) ether of 14 in the presence of TBS ether was unsuccessful. Thus, TBAF-mediated cleavage of the TBS ether of 14 followed by MOM-deprotection and global

TBS protection gave rise to disaccharide disulfide acceptor 5. Finally, LiDBB-mediated reductive lithiation of the *L*-rhodinosyl phenylsulfide of 6^{13a} at −78 °C afforded the corresponding axial glycosyl lithium which then reacted with asymmetric disulfide acceptor 5 to give the desired trisaccharide donor 3 in 78% yield.

With trisaccharide donor 3 in hand, we turned our attention to the synthesis of trisaccharide-derived asymmetric disulfide acceptor 4. As shown in Scheme 3, the alcohol of known *L*-

Scheme 3. Attempted Synthesis of S-Linked Hexasaccharide (2)

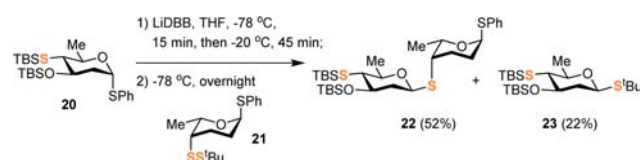


amicetosyl phenylsulfide 15¹⁷ underwent triflation and subsequent S_N2 substitution with cesium thioacetate to furnish thioacetate 16. Reductive removal of the S-acetyl group followed by silylation of the resulting free thiol afforded donor 7. Likewise, our umpolung-based α-S-glycosylation¹³ between donor 7 and disaccharide acceptor 5 furnished S-linked trisaccharide 17 in 76% yield. Next, selective deprotection of triisopropylsilyl thioether 17 in the presence of TBS ether led to the formation of the corresponding free thiol which was then converted to the trisaccharide asymmetric disulfide acceptor 4 in 56% yield over two steps. In order to prepare the target hexasaccharide 2, reductive lithiation of trisaccharide donor 3 followed by anomerization (−30 °C)¹³ afforded the corresponding equatorial glycosyl lithium which was subjected to the reaction with trisaccharide asymmetric disulfide acceptor 4 at −78 °C. However, it was very disappointing to find out that this 3 + 3 coupling did not provide the desired S-linked hexasaccharide 18. Rather, a trisaccharide-derived β-*tert*-butylthioglycoside 19 was obtained in approximately 5% yield. Mechanistically, compound 19 may be formed by nucleophilic attack of the trisaccharide donor 3-derived equatorial glycosyl lithium to the sulfur atom next to the *tert*-butyl group of disulfide acceptor 4. In addition, we noticed that significant amounts of trisaccharide disulfide acceptor 4 were recovered, which indicated the low reactivity of trisaccharide disulfide acceptor 4 probably due to the steric bulkiness (the *tert*-butyl disulfide moiety is *cis*- to the C6-methyl group of the nonreducing sugar moiety of 4).

We speculated that the reactivity of the disulfide acceptor might be enhanced by truncating the trisaccharide 4 to a

monosaccharide (cf. **21**, Scheme 4). In order to test this idea, we prepared monosaccharide donor **20** and monosaccharide-

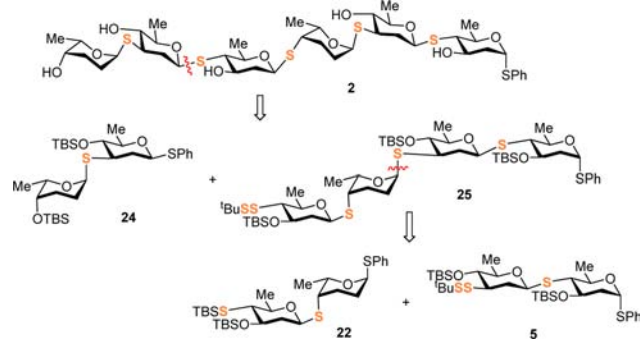
Scheme 4. Synthesis of S-Linked Disaccharide **22**



derived asymmetric disulfide acceptor **21**.¹⁸ Gratifyingly, reductive lithiation of donor **20** followed by anomerization ($-20\text{ }^{\circ}\text{C}$)¹³ afforded the corresponding equatorial glycosyl lithium which subsequently reacted with disulfide acceptor **21** to give desired β -S-linked disaccharide **22** in 52% yield together with β -*tert*-butylthioglycoside **23** (22% yield). Disaccharide **22** corresponds to the disaccharide fragment CD of hexasaccharide **2** (cf. Figure 1).

With successful preparation of β -S-linked disaccharide **22**, we revised our synthetic strategy to that shown in Scheme 5. Thus,

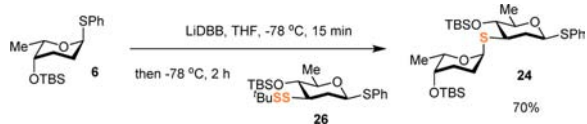
Scheme 5. Revised Retrosynthesis of S-Linked Hexasaccharide of Landomycin A (**2**)



S-linked hexasaccharide **2** could potentially be obtained by **2** + **4** coupling of disaccharide donor **24** and tetrasaccharide-derived asymmetric disulfide acceptor **25** via our reported umpolung S-glycosylation.¹³ In turn, tetrasaccharide-derived asymmetric disulfide acceptor **25** might be obtainable from the previously prepared disaccharide donor **22** and disaccharide-derived disulfide **5**.

As shown in Scheme 6, α -S-disaccharide donor **24** can be readily prepared in 70% yield by reductive lithiation of **L**-

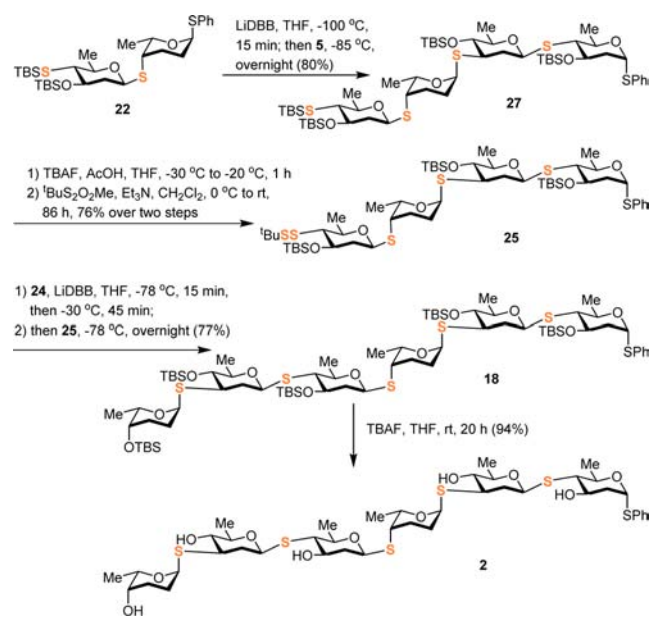
Scheme 6. Synthesis of Disaccharide Donor **24**



rhodosyl phenylsulfide **6**^{13a} at $-78\text{ }^{\circ}\text{C}$ followed by reaction with asymmetric disulfide **26**.¹⁹ Disaccharide **24** corresponds to the disaccharide fragment EF of hexasaccharide **2** (cf. Figure 1).

With disaccharide donors **22** and **24** as well as disaccharide disulfide acceptor **5** (fragment AB) available, we executed the final stages of our synthesis of S-linked hexasaccharide **2** (Scheme 7). Accordingly, reductive lithiation of disaccharide-derived phenylsulfide **22** at $-100\text{ }^{\circ}\text{C}$ gave the corresponding axial glycosyl lithium which then reacted with disaccharide-

Scheme 7. Revised Retrosynthesis of S-Linked Hexasaccharide of Landomycin A (**2**)



derived asymmetric disulfide **5** at $-85\text{ }^{\circ}\text{C}$ to afford desired tetrasaccharide **27** in 80% yield (fragment ABCD). Selective deprotection of the *tert*-butyldimethylsilyl thioether of **27** in the presence of three TBS ethers led to the formation of the free thiol which reacted with *S*-*tert*-butylmethanethiosulfonate to afford asymmetric disulfide acceptor **25**. Similarly, LiDBB-mediated reductive lithiation of the disaccharide-derived donor **24** followed by anomerization ($-30\text{ }^{\circ}\text{C}$)¹³ afforded the corresponding equatorial lithium which then reacted with the asymmetric disulfide acceptor **25** to give the desired fully protected β -S-linked hexasaccharide **18** in 77% yield. Finally, global deprotection of five *tert*-butyldimethylsilyl ethers using TBAF at room temperature afforded desired S-linked hexasaccharide **2**.

In conclusion, we have described the first stereoselective synthesis of an S-linked hexasaccharide subunit of landomycin A. This synthesis has been achieved using our previously reported umpolung reactivity-based S-glycosylation via sulfenylation of stereochemically defined glycosyl lithium species with asymmetric sugar-derived disulfides. Preparation of analogs of landomycin A bearing S-linked 2-deoxy sugar subunits and their biological evaluation is underway and will be reported in due course.

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.orglett.5b02223.

Experimental procedure; characterization data for all new compounds (PDF)

■ AUTHOR INFORMATION

Corresponding Author

*E-mail: Jianglong.Zhu@Utoledo.edu.

Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

We are grateful to the National Science Foundation (CHE-1213352), Ohio Cancer Research Associates, and The University of Toledo for supporting this research. We thank Mr. Isaac Morales (University of Toledo), Ms. Mikayla Becker (Maumee High School, Maumme, OH), and Cristin Reno (Southview High School, Sylvania, OH) for experimental assistance.

■ REFERENCES

- (1) (a) Henkel, T.; Rohr, J.; Beale, J. M.; Schwenen, L. *J. Antibiot.* **1990**, *43*, 492–503. (b) Shaaban, K. A.; Srinivasan, S.; Kumar, R.; Damodaran, C.; Rohr, J. *J. Nat. Prod.* **2011**, *74*, 2–11. (c) Shaaban, K. A.; Statatkin, C.; Damodaran, C.; Rohr, J. *J. Antibiot.* **2011**, *64*, 141–150.
- (2) (a) Depenbrock, H.; Bornschlegl, S.; Peter, R.; Rohr, J.; Schmid, P.; Schweighart, P.; Block, T.; Rastetter, J.; Hanauske, A.-R. *Ann. Hematol.* **1996**, *73*, A80/316. (b) Rohr, J.; Wohlert, S.-E.; Oelkers, C.; Kirschning, A.; Ries, M. *Chem. Commun.* **1997**, 973–974.
- (3) Crow, R. T.; Rosenbaum, B.; Smith, R., III; Guo, Y.; Ramos, K. S.; Sulikowski, G. A. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 1663–1666.
- (4) Weber, S.; Zolke, C.; Rohr, J.; Beale, J. M. *J. Org. Chem.* **1994**, *59*, 4211–4214.
- (5) Roush, W. R.; Neitz, R. J. *J. Org. Chem.* **2004**, *69*, 4906–4912.
- (6) Yang, X.-Y.; Fu, B.-Q.; Yu, B. *J. Am. Chem. Soc.* **2011**, *133*, 12433–12435.
- (7) Guo, Y.; Sulikowski, G. A. *J. Am. Chem. Soc.* **1998**, *120*, 1392–1397.
- (8) Roush, W. R.; Bennett, C. E. *J. Am. Chem. Soc.* **2000**, *122*, 6124–6125.
- (9) Yu, B.; Wang, P. *Org. Lett.* **2002**, *4*, 1919–1922.
- (10) Kirschning, A. *Eur. J. Org. Chem.* **1998**, *1998*, 2267–2274.
- (11) Zhou, M.; O'Doherty, G. A. *Org. Lett.* **2008**, *10*, 2283–2286.
- (12) Tanaka, H.; Yamaguchi, S.; Yoshizawa, A.; Takagi, M.; Shin-ya, K.; Takahashi, T. *Chem. - Asian J.* **2010**, *5*, 1407–1424.
- (13) (a) Baryal, K. N.; Zhu, D.; Li, X.; Zhu, J. *Angew. Chem., Int. Ed.* **2013**, *52*, 8012–8016. (b) Baryal, K. N.; Zhu, J. *Synlett* **2014**, *25*, 308–312.
- (14) For additional references on the stereoselective synthesis of S-linked 2-deoxy glycosides, see: (a) Crich, D.; Ritchie, T. J. *Carbohydr. Res.* **1989**, *190*, C3–C6. (b) Sherry, B. D.; Loy, R. N.; Toste, F. D. *J. Am. Chem. Soc.* **2004**, *126*, 4510–4511. (c) Issa, J. P.; Lloyd, D.; Steliotes, E.; Bennett, C. S. *Org. Lett.* **2013**, *15*, 4170–4173.
- (15) For reviews on the synthesis and application of S-linked glycosides (thioglycosides), see: (a) Driguez, H. *Top. Curr. Chem.* **1997**, *187*, 85–116. (b) Szilagyi, L.; Varela, O. *Curr. Org. Chem.* **2006**, *10*, 1745–1770. (c) Driguez, H. *ChemBioChem* **2001**, *2*, 311–318.
- (16) Koo, B.; McDonald, F. E. *Org. Lett.* **2007**, *9*, 1737–1740.
- (17) Synthesis of compound **15** was previously reported; see: Kahne, D. E.; Goodnow Jr, R. A.; Taylor, C. M.; Yan, L. U.S. Patent 5,700,916, 1997. In addition, we have described the details for the preparation and characterization of compound **15** in the [Supporting Information](#).
- (18) See [Supporting Information](#) for the preparation of donor **20** and acceptor **21**.
- (19) See [Supporting Information](#) for the preparation of disulfide acceptor **26**.